

FLAVONOIDS OF *Campanula alata* AND THEIR ANTIOXIDANT ACTIVITY

Ouassila Touafek,¹ Zahia Kabouche,^{1*} Ignacio Brouard,²
and Jaime Barrera Bermejo²

UDC 547.972

In addition to their use as ornamental plants, *Campanula* (Campanulaceae) species are used in traditional medicine to treat various diseases such as tonsillitis, laryngitis, bronchitis, and warts [1]. They possess stimulant properties and are used as emetics; they also possess refreshing, antiallergic, antiphlogistic, antioxidant, spasmolytic, antiviral, and antimicrobial properties [2, 3]. Flavonoids are the most important secondary metabolites of the *Campanula* genus [4–9].

Campanula alata Desf. (Campanulaceae) is an endemic species [10] collected from Constantine (Eastern Algeria) in April 2004 and authenticated by Prof. Gerard De Belair (Annaba, Algeria).

A voucher specimen was deposited in the Herbarium of the Laboratory of Therapeutic Substances (LOST) at Mentouri University (LOST/Ca/04/04).

Air-dried and powdered aerial parts (1 kg) of *Campanula alata* Desf. were macerated in a methanolic solution (70%). The extract was successively concentrated to dryness (under low pressure); the residue was dissolved in boiling water and extracted with petroleum ether, dichloromethane, ethyl acetate, and *n*-butanol, successively.

The butanolic extract was column chromatographed on Polyamid SC6 eluted with toluene–methanol with increasing polarity. Whatman 3MM paper chromatography using 15% AcOH and BAW [*n*-BuOH–AcOH–H₂O, 4:1:5 (upper phase)] and TLC on Polyamid DC6 eluted with H₂O–MeOH–metylethylketone–acetylacetone (13:3:3:1) followed by column flash chromatography over Sephadex LH-20 in MeOH led to four pure flavonoids **1–4** and a caffeic ester (**5**), which were identified by using UV, ¹H NMR, ¹³C NMR, and MS analysis [11–13].

Acid Hydrolysis. The pure compounds were treated with 2 M HCl at 100°C for 1 h. The hydrolysates were extracted with EtOAc, and the aglycones were identified by their UV spectra in methanol and by comparison of their *R_f* with authentic samples.

Sugars were identified in the aqueous residue by comparison with authentic samples on silica gel TLC impregnated with 0.2 M NaH₂PO₄ [solvent Me₂CO–H₂O (9:1)] and revealed with aniline malonate.

Antioxidant Activity. The radical scavenging activity of the *n*-butanolic extract of *Campanula alata* (BECA) was measured by the slightly modified method of Hatano [14]. One milliliter of a 0.2 mM DPPH methanol solution was added to 4 mL of various concentrations of the extract in methanol. The mixture was shaken vigorously and left to stand at room temperature. After 30 min, the absorbance of the solution was measured at 517 nm and the antioxidant activity calculated using the following equation: Scavenging capacity % = 100 – [(Ab of sample – Ab of blank) × 100/Ab of control]. Methanol (1 mL) plus plant extract solution (4 mL) were used as a blank, while DPPH solution plus methanol was used as a negative control. The positive control was DPPH solution plus 1 mM rutin. Extract concentration providing 50% inhibition (IC₅₀) was calculated from the plot of inhibition percentage against extract concentration.

Compound 1. C₁₅H₁₀O₇, yellow needles (acetone), mp >300°C. UV (MeOH, λ_{max}, nm): 256, 374; +AlCl₃: 271, 451; +AlCl₃/HCl: 266, 302sh, 361sh, 427. ¹H NMR spectrum (400 MHz, acetone-d₆, δ, ppm, J/Hz): 7.81 (1H, br.s, H-2'), 7.70 (1H, d, J = 8.0, H-6'), 6.99 (1H, d, J = 8.0, H-5'), 6.53 (1H, br.s, H-8), 6.27 (1H, br.s, H-6). This compound was characterized as quercetin [15].

1) Laboratoire d'Obtention de Substances Therapeutiques (L.O.S.T), Faculte des Sciences Exactes, Universite Mentouri-Constantine, Campus Chaabat Ersas, 25000 Constantine, Algerie, e-mail: zkabouche@yahoo.com; 2) Instituto de Productos Naturales y Agrobiologia-C.S.I.C.-Instituto Universitario de Bio-Organica “Antonio Gonzalez”, Universidad de La Laguna, Av. Astrofisico F. Sanchez 3, 38206 La Laguna, Tenerife, Spain. Published in Khimiya Prirodnykh Soedinenii, No. 6, pp. 825–826, November–December, 2010. Original article submitted July 15, 2009.

Compound 2. $C_{21}H_{20}O_{11}$, mp 239–242°C (MeOH). UV (MeOH, λ_{max} , nm): 267, 336; +NaOH: 275, 405; +AlCl₃: 275, 291, 330, 423; +AlCl₃/HCl: 275, 296, 355, 385; +NaOAc: 267, 400; +NaOAc/H₃BO₃: 260, 370. ¹H NMR (250 MHz, CD₃OD, δ, ppm, J/Hz): 7.42 (1H, dd, J = 9.4, J = 2.2, H-6'), 7.40 (1H, d, J = 2.2, H-2'), 6.93 (1H, d, J = 9.4, H-5'), 6.81 (1H, d, J = 2.2, H-8), 6.63 (1H, s, H-3), 6.52 (1H, d, J = 2.2, H-6), 5.10 (1H, d, J = 10, H-1''), 3.25–4.25 (sugar protons). Acid hydrolysis of compound 2 produced luteolin and glucose. This compound was characterized as 5,3',4'-trihydroxy-7-O-glucosylflavone (luteolin 7-glucoside) [16].

Compound 3. Mp 218–212°C. UV (MeOH, λ_{max} , nm): 257, 353; + NaOH: 272, 331, 404; + AlCl₃: 273, 300 sh, 434; +AlCl₃/HCl: 269, 359 sh; 396. ¹H NMR (400 MHz, DMSO-d₆, δ, ppm, J/Hz): 7.60 (1H, d, J = 2.3, H-2'), 7.60 (1H, m, H-6'), 6.85 (1H, d, J = 8.9, H-5'), 6.40 (1H, d, J = 1.9, H-8), 6.20 (1H, d, J = 1.9, H-6), 5.40 (1H, d, J = 6.9, H-1''Glc), 3.2–4.9 (sugar protons).

Acid hydrolysis of 3 produced quercetin and D-glucose. Compound 3 was identified as quercetin 3-O-glucoside [17].

Compound 4. $C_{27}H_{30}O_{16}$, mp 250–254°C. UV (MeOH, λ_{max} , nm): 257, 300sh, 356, +NaOH: 272, 325, 407; + AlCl₃: 275, 290, 350sh; +HCl: 268, 285sh, 350, 390; +NaOAc: 271, 385; +H₃BO₃: 263, 378. ¹H NMR (250 MHz, CD₃OD, δ, ppm, J/Hz): 7.80 (1H, d, J = 2, H-2'), 7.75 (1H, dd, J = 9, J = 2, H-6'), 6.85 (1H, d, J = 9, H-5'), 6.3 (1H, d, J = 2, H-8), 6.20 (1H, d, J = 2, H-6), 5.12 (1H, d, J = 7, H-1''Glc), 4.55 (1H, d, H-1'''Rha), 1.1 (3H, d, J = 6.2, H-6'''Rha), 3.20–3.90 (10H, protons of rutinose). ¹³C NMR (62.89 MHz, CD₃OD, δ): 177.9 (C-4), 164.6 (C-7), 161.5 (C-5), 157.9 (C-2), 157.06 (C-9), 148.4 (C-4'), 144.4 (C-3'), 134.2 (C-3), 122.1 (C-6'), 121.6 (C-1'), 116.2 (C-5'), 114.6 (C-2'), 104.17 (C-10), 103.3 (C-1''), 100.9 (C-1'''), 98.5 (C-6), 93.4 (C-8), 76.7 (C-5''), 75.7 (C-3''), 74.3 (C-2''), 72.5 (C-4''), 70.7 (C-4''), 70.6 (C-2''), 69.9 (C-3''), 68.3 (C-5''), 67.1 (C-6''), 64.4 (C-6''). Identified as quercetin-3-O-rutinoside [18].

Compound 5. $C_{10}H_{10}O_4$, yellowish crystals, mp 162–163°C. IR (KBr, v, cm⁻¹): 3493 (OH), 3308 (C-H aromatic), 1681 (C=O), 1603 (C=C aromatic). UV (MeOH, λ_{max} , nm): 328, 300, 244, 219. ¹H NMR (400 MHz, CD₃OD, δ, ppm, J/Hz): 7.56 (d, J = 16.5, H-7), 7.05 (d, J = 2, H-2), 6.96 (dd, J = 8 and 2, H-6), 6.80 (d, J = 8, H-5), 6.27 (d, J = 16.5, H-8), 3.77 (s, OCH₃). ¹³C NMR (100 MHz, CD₃OD, δ): 113.6 (C-8), 113.9 (C-2), 115.3 (C-5), 121.8 (C-6), 126.5 (C-1), 145.7 (C-3), 145.8 (C-7), 148.4 (C-4), 168.6 (C-9). Identified as methyl caffeoate [19].

The butanolic extract of *Campanula alata* (BECA) exhibited good activity (IC_{50} 25.77 ± 0.2 μg/mL) compared with rutin (IC_{50} 3.01 ± 0.2 μg/mL).

ACKNOWLEDGMENT

The authors thank the DG/RSDT, MESRS, Algeria for financial support.

REFERENCES

1. J. Roi, *Traite des Plantes Medicinales Chinoises*, Encyclopedie biologique, Paris, 1955.
2. J. F. Morton, *Major Medicinal Plants, Botany, Culture and Uses*, Charles C. Thomas Publisher, Springfield, U.S.A., 1977.
3. J. C. Rameau, D. Mansion, and G. Dume, *Flore Forestiere Francaise*, Guide Ecologique illustre, 1, Paris, 1989.
4. L. S. Teslov and L. N. Koretskaya, *Khim. Prir. Soedin.*, 786 (1983)
5. L. S. Teslov, *Khim. Prir. Soedin.*, 790 (1984).
6. L. S. Teslov, *Khim. Prir. Soedin.*, 593 (1988).
7. L. S. Teslov, *Khim. Prir. Soedin.*, 271 (1990).
8. K. Brandt, T. Kondo, H. Aoki, and T. Goto, *Phytochemistry*, **33**, 209 (1993).
9. M. U. Dumlu, E. Gurkan, and E. Tuzlaci, *Nat. Prod. Res., Part A: Struct. Synth.*, **22**, 477 (2008).
10. P. Quezel and S. Santa, *Nouvelle Flore de l'Algérie et des Régions Désertiques et Méridionales*, Tome II, Editions CNRS, Paris, 1963.
11. K. R. Markham, *Techniques of Flavonoid Identification*, Academic Press, London, 1982.

12. K. R. Markham and H. Geiger, *¹H Nuclear Magnetic Resonance Spectroscopy of Flavonoids and Their Glycosides in Hexadeuterodimethyl Sulfoxide*, in: *The Flavonoids. Advances in Research Since 1986*, J. B. Harborne (ed.), Chapman and Hall, 1993.
13. K. R. Markham and H. Geiger, *¹H NMR Spectroscopy of Flavonoids and Their Glycosides in Hexadeuterodimethylsulfoxide*, in: *The Flavonoids*, J. B. Harborne (ed.), 1993, Chapman and Hall, London, 1994.
14. T. Hatano, H. Kagawa, T. Yasuhara, and T. Okuda, *Chem. Pharm. Bull.*, **36**, 2090 (1988).
15. E. S. Nedel'ko and G. K. Nikonov, *Chem. Nat. Comp.*, **23**, 254 (1987).
16. V. V. Stetskov and V. L. Shelyuto, *Chem. Nat. Comp.*, **18**, 493 (1982).
17. A. I. Syrchina, V. G. Gorokhova, N. A. Tyukavkina, V. A. Babkin, and M. G. Voronkov, *Chem. Nat. Comp.*, **16**, 245 (1980).
18. N. Navrezova, M. Agzamova, N. N. Stepanichenko, and B. Makhsudova, *Chem. Nat. Comp.*, **22**, 229 (1986).
19. S. K. Inayama, H. Karimaya, T. Hori, T. Ohkura, M. Kawamata, and T. Hikichi, *Chem. Pharm. Bull.*, **32**, 1135 (1984).